Organic & Biomolecular **Chemistry**

www.rsc.org/obc Volume 10 | Number 14 | 14 April 2012 | Pages 2701–2896

ISSN 1477-0520

RSCPublishing

PERSPECTIVE Mats Larhed et al.

Microwave-assisted synthesis of small molecules targeting the infectious diseases tuberculosis, HIV/AIDS, malaria and hepatitis C

1477-0520(2012)10:14;1-G

Organic & Chemistry

 C ito this: Ora, Piomo Cite this: *Org. Biomol. Chem.,* 2012, **10**, 2713

Microwave-assisted synthesis of small molecules targeting the infectious diseases tuberculosis, HIV/AIDS, malaria and hepatitis C

Johan Gising, Luke R. Odell and Mats Larhed*

Received 1st November 2011, Accepted 1st December 2011 DOI: 10.1039/c2ob06833h

The unique properties of microwave in situ heating offer unparalleled opportunities for medicinal chemists to speed up lead optimisation processes in early drug discovery. The technology is ideal for small-scale discovery chemistry because it allows full reaction control, short reaction times, high safety and rapid feedback. To illustrate these advantages, we herein describe applications and approaches in the synthesis of small molecules to combat four of the most prevalent infectious diseases; tuberculosis, HIV/ AIDS, malaria and hepatitis C, using dedicated microwave instrumentation. **Bownloaded Community at Albany on 24 March 2012**
 DERSPECTIVE
 **Microwave-assisted synthesis of small molecules targeting the infectious diseases tuberculosis, HIV/AIDS, malaria and hepatitis C

Microwave-assisted syn**

1. Introduction

Microwave ovens were introduced into analytical chemistry in the late 1970s and the first articles reporting that microwaves could be used to accelerate organic reactions appeared in 1986 .^{1,2} The uptake of this new heating technology in the synthetic chemistry community was initially slow due to problems with reproducibility, controllability, and safety aspects, together with a generally low degree of understanding of the basic principles of microwave (MW) heating (the fact that all polar

Organic Pharmaceutical Chemistry, Department of Medicinal Chemistry, Uppsala Biomedical Centre, Uppsala University, Box 574, SE-751 23 Uppsala, Sweden. E-mail: mats.larhed@orgfarm.uu.se; Fax: +46 18 4714474; Tel: +46 18 4714667

solvents can be heated with microwaves and not only water was not generally recognised). $3-7$ However, it has gained in popularity, and sophisticated microwave systems can today be found in most well-equipped organic and medicinal chemistry laboratories in industry and academia.⁸ The reason behind its popularity lies in the user friendly instrumentation and its outstanding ability to reduce reaction times by superheating of organic reactions in sealed vessels.

All domestic microwave ovens, as well as the commercial reactors found in chemistry laboratories, operate at 2.45 GHz in order not to interfere with the wavelengths used for radar or telecommunication. This frequency corresponds to a wavelength of 12.2 cm and is located between the infrared and radiowave wavelengths in the electromagnetic spectrum. Dipolar polarisation is

Johan Gising

Johan Gising was born in 1981 in Falun, Sweden. In 2001, he commenced undergraduate studies at Uppsala University and, in 2006, he received a master degree in pharmaceutical chemistry. In the same year he began his PhD studies under Professor Mats Larhed and Professor Anders Karlén and he will defend his thesis during the spring of 2012. His research focus has been the design, synthesis and physio-

chemical properties of new inhibitors targeting enzymes for the potential treatment of tuberculosis, hepatitis C and HIV.

Luke Odell

graduated with an Honours BSc in Forensic Science from the University of Newcastle, Australia in 2002. He completed his PhD studies at the same university under the guidance of Professor Adam McCluskey in 2006 working on the synthesis of enzyme inhibitors. In 2006, he took up a postdoctoral position with Professor Mats Larhed at Uppsala

Luke Odell was born in Tamworth, Australia in 1981. He

University. Since 2009, he has been an Associate Professor at Uppsala University and his research interests include microwave-assisted metal catalysis, heterocyclic chemistry and medicinal chemistry.

the main mechanism accounting for microwave heating in polar systems although in materials containing ions, ionic conduction starts to prevail.³ In contrast to classical heating, microwaves generate heat inside the bulk of the sample (in situ heating) and will be distributed from inside and out (no wall heating) causing the sample to heat up evenly and rapidly. It is important to realise that, in principle, all reactions that are driven by heat can be performed with microwave heating. In theory, the use of nonpolar microwave-transparent solvents should give poor heating but in reality the presence of polar or ionic starting materials, reagents, catalysts and additives in the reaction system enables satisfactory heating of most laboratory-scale organic reactions. By using sealed reaction vessels it is thus possible to rapidly and safely heat nearly all reaction systems and to maintain the temperature much above the atmospheric reflux temperature.⁹ Download mechanism accounting for microwave heuting in polar well-defined regions of maxima and minima field systems at Albany interaction (and the control in a control in a control in a control in a control in the control

Most of the examples of microwave-assisted organic synthesis before 2002 were carried out using domestic microwave ovens.⁴ These ovens produce a non-continuous heating pattern within the microwave cavity due to an on–off cycle of the power supply, leading to problems associated with reproducibility and the accuracy of (or lack of) temperature measurements. Furthermore, domestic microwave ovens are not constructed to withstand an explosion and are also associated with other safety hazards. To solve the problems mentioned above, modern, scientific microwave reactors for organic chemistry applications were developed in the mid to late 1990s and today there are several models available on the market.

Two general classes of dedicated microwave reactors are currently available, multimode and single-mode.^{4,5} In the multimode instruments, microwaves are randomly distributed within the large cavity and less defined regions of high and low intensity are produced. These instruments often rely on continuous rotation of the samples in the cavity to ensure an even energy distribution. This is a design suitable for parallel or large-scale (<1000 mL) processing. On the other hand, in a single-mode microwave cavity a continuous standing wave is generated with

Mats Larhed

Mats Larhed received his PhD in 1997 and became a full professor in 2007. He holds the position as Chair of the Division of Organic Pharmaceutical Chemistry and as Head of Preclinical PET at Uppsala University. Dr Larhed's main research focus has been towards the development of fast, selective and robust synthetic methods for use in preparative medicinal chemistry. His work in microwave-assisted

metal-catalysis covers different types of palladium-catalyzed coupling reactions, gas-free carbonylations and the development of environmentally benign chemical transformations. During the last ten years he has been increasingly engaged in the development of enzyme inhibitors for potential treatment of HIV, malaria, Alzheimer's disease and tuberculosis.

well-defined regions of maxima and minima field strengths.¹⁰ The single-mode equipment is more energy efficient and allows the placement of the reaction vessel at a fixed position with high energy density. The temperature level is varied by software-controlled modulation of the continuous-wave output using an IRsensor or a fibre-optic probe. As IR sensors actually measure the temperature of the reaction vessel walls, they will not always reflect the temperature inside the vessel. Therefore multiple fibreoptic sensors immersed in the reaction mixture should be used if a correct temperature measurement is required. 11 Importantly, the difficulties concerning correct temperature measurements are also true for multimode reactors and especially in case of parallel processing of different reaction mixtures.¹²

The use of single-mode cavities is of particular importance in the acceleration of sealed small-scale (0.2–20 mL) reactions requiring high temperature, pressure monitoring and safe processing, making the equipment ideal for reaction scouting and chemical optimisation. 13 Further, automated, sequential singlemode reactors with disposable reaction vials are of huge importance for compound production in discovery projects.¹⁴

While the commercially available microwave equipment of today is very useful in lead generation and lead optimisation, it is necessary to obtain larger quantities of compound at the later stages of development. However, the invention of microwave flow reactors, $3,15-19$ or the use of high quality large-scale multimode batch reactors²⁰ may provide attractive microwave solutions to the problem of scalability.

2. Infectious diseases

Infectious diseases are a major cause of death worldwide claiming the lives of as many as 15 million people each year and, to make matters worse, people carrying one infectious disease become more susceptible to other diseases. 21 Accordingly, significant effort has been put into identifying new and effective ways to combat these diseases. Moreover, many of the pathogens exhibit extremely fast mutation rates, leading to the emergence of drug resistance and in some cases, extreme drug resistance. This has resulted in the need for the continuous development of new therapeutic agents and effective combination therapies. This perspective will focus on the use of single-mode microwave irradiation for the synthesis of small molecules in laboratory scale for the potential treatment of tuberculosis, HIV/ AIDS, malaria and hepatitis C; four of the most dangerous and prevalent infectious diseases.

2.1 Tuberculosis

One of the world's deadliest diseases is tuberculosis (TB) which is caused by infection with Mycobacterium tuberculosis (MTB). Today, 2 billion people or one third of the world's population are infected with the TB bacillus and each year over 9 million new TB cases are discovered.²² In 2009, 1.7 million people died from TB which equates to over 4600 deaths a day.²² The emergence of multidrug-resistant TB (MDR-TB) and extensively drug-resistant TB (XDR-TB) confirms the urgent need for development of new TB specific drugs.^{23,24}

MTB glutamine synthethase (MTB-GS) is believed to be involved in cell wall biosynthesis $\frac{3}{5}$ and the pathogens ability to

Scheme 1 Synthesis of MTB-GS inhibitors via a microwave-assisted palladium-catalysed alkoxycarbonylation.

inhibit phagosome-lysosome fusion and phagosome acidification.^{26,27} An inhibitor of the enzyme, L-methionine- (SR) -sulfoximine (MSO, see Scheme 1), has been shown to hinder bacterial growth both in vitro²⁸ and in vivo²⁹ confirming MTB-GS as a promising and druggable target.

Recently Gising and co-workers reported the synthesis and biological evaluation of tri-substituted imidazoles as a new class of MTB-GS inhibitors (Scheme 1).³⁰ Through a palladium-catalysed alkoxycarbonylation, 2-bromo-6-methoxynaphthalene was coupled with ethanol utilising single-mode microwave irradiation as heating source to give the ethyl ester 1 in excellent yield (93%). A sluggish S_NAr substitution of the fluorine with methanamine gave the final inhibitor 2. Compound 2 was 10 fold more active towards MTB-GS than MSO. In addition, an analogue to 2 was the most potent compound in the series $(IC_{50} =$ 0.049 μM).

In 2009, Odell et al. investigated functionalised 3-aminoimidazo[1,2-a]pyridines as MTB-GS inhibitors derived from a HTS.³¹ The target compounds were synthesised in moderate to good yields via an Ugi-type, multi-component heterocylisation from a 2-aminopyridine, an aldehyde and an isocyanate (Scheme 2). Compound 3 displayed considerable better activity *in vitro* than MSO (MTB-GS $IC_{50} = 0.38 \mu M$ *vs.* 51 μ M).

For an additional multi-component, microwave-assisted, heterocylisation in the synthesis of isoniazid analogues see the recent report by Manjashetty et al.³²

Isopentyl diphosphate, the precursor to a group of essential isoprenoids for the TB bacilli is synthesised through the nonmevalonate pathway.³³ In eukaryotes however, isopentyl diphosphate is instead produced in the classical mevalonate pathway. As the routes differ from man to bacilli, the non-mevalonate

Scheme 2 Microwave-assisted synthesis of MTB-GS inhibitors via multi-component heterocyclisation.

fosmidomycin

Scheme 3 Synthesis of MTB-DXR inhibitors via a microwave promoted oxidative Heck reaction.

pathway enzymes are suitable targets for drug discovery. MTB 1-deoxy-D-xylulose 5-phosphate reductoisomerase (MTB-DXR, also referred to as IspC) is involved in the non-mevalonate pathway³⁴ and fosmidomycin³⁵ (see Scheme 3) is a known inhibitor of the enzyme. Furthermore, knockouts of the DXR gene in E. coli. are lethal promoting DXR as promising target for drug development.^{36,37}

In the exploration of fosmidomycin analogues for MTB-DXR inihibition, Andaloussi et al. introduced various aromatic groups α to the phosphonic acid (Scheme 3).³⁸ An oxidative Heck reaction under single-mode microwave heating was employed to

substitute the boronic acid for an acrylaldehyde using palladium (I) acetate and 2,9-dimethyl-1,10-phenanthroline (dmphen) as the catalytic system.39,40 The cinnamalaldehyde 4 allowed easy access to the final compounds, exemplified by 5, one of the most active compounds. Compound 5 displayed a comparable anti-TB activity to fosmidomycin.

In related studies, the same group explored the aromatic substituents further 40 and also investigated bioisosteres to the phosphonic and hydroxamic acid groups.⁴¹

MTB protein tyrosine phosphatase (MptpB) is an important signalling enzyme in the bacteria. A mutant strain of MTB with disrupted MptpB activity impairs the bacterial survival in a guinea pig model.⁴² The complete function of the enzyme is not yet fully understood and therefore the discovery of inhibitors of the enzyme would be of great value. Finding selective inhibitors is a challenge as the active site is highly conserved by most members in the protein tyrosine phosphatase family.⁴³

In an effort to identify selective MptpB inhibitors, a library of ∼3500 bidentate compounds was designed by Tan and coworkers (Scheme 4).⁴⁴ In the search for additional interactions to an allosteric site, a linker was incorporated between the acidic active site binder and the anthracene moiety. In the synthesis of one of the active site binders, a palladium-catalysed Negishi cross-coupling was performed under temperature controlled microwave heating giving 6 in good yield. In the final step, 11 alkynes containing active site binders were reacted with numerous azide linkers to form triazoles.^{45,46} Compound 7 displayed submicromolar activity towards MptpB and was one of the most promising entities in the 3500 compound library.

For the microwave promoted synthesis of other MTB inhibitors through metal catalysis see Kim *et al.*⁴⁷ and Lagerlund et al.⁴⁸

In a report from 2010, Manna et al. described the synthesis of MTB and MDR-TB inhibitors through heterocyclisation to give 1,3,5-trisubstituted-4,5-dihydro-1H-pyrazoles.⁴⁹ The yield for the cyclisation was improved by applying single-mode microwave heating in sealed vessels and reaction times could be decreased to 12–22 min instead of reflux for 6–10 h. A set of 14 α,β-unsaturated ketones were reacted with two hydrazides to

Scheme 4 Synthesis of MptpB inhibitors using a microwave-assisted Negishi cross-coupling.

Scheme 5 Synthesis of MTB inhibitors via a microwave heated 5membered heterocyclisation.

Scheme 6 Synthesis of MTB inhibitors via a microwave-assisted 6membered heterocyclisation.

yield the final products in good yields (Scheme 5). Of the 28 1,3,5-trisubstituted-4,5-dihydro-1H-pyrazoles synthesised, inhibitor 8 was amongst the most active compounds and it was twice as active against MDR-TB than isoniazid. Furthermore, compound 8 was reported to lower the Mycobacterium tuberculosis count in lung and spleen two log units further than isoniazid in a mouse model.

A number of related examples of microwave-assisted, 5-membered heterocyclisations towards MTB inhibitor synthesis have also been reported.31,50–⁵⁵

Zhou *et al.* synthesised a library of pyrano-annulated 5,6,7,8tetrahydro-1,6-naphthyridines, exemplified by 9 (Scheme 6), in the search for active compounds towards MTB.⁵⁶ The

Scheme 7 Microwave-assisted synthesis of MTB inhibitors via a condensation reaction.

intramolecular cobalt-catalysed $[2 + 2 + 2]$ cyclisation gave 9 in excellent yield under microwave irradiation for 30 min at 180 °C. For diversity, four variants of 9 were synthesised where $R¹$ and $R²$ were altered with alkyls and aryls. After PMB-deprotection the four resulting secondary amines were coupled with a diverse set of 8 isocyanates, acyl chlorides and sulfonyl chlorides to yield ureas, amides and sulphonamides, respectively. In the constructed library of 96 compounds, the scaffold with $R¹$ = 2-methyl-4-methoxyphenyl and R^2 = phenyl was shown to be most promising as low micromolar inhibitors were found in all three N-2 derivatised classes, exemplified by 10 in Scheme 6. The sulfonamide 10 was the most promising compound in the library with a low micromolar MIC (minimum inhibitory concentration) and a good cytotoxicity profile.

Synthesis of other microwave-assisted 6-membered heterocyclisations towards MTB inhibitors have been reported by Balamurugan et al.,⁵⁷ Kim et al.⁵⁸ and Manjashetty et al.³²

In 2008, Biava and co-workers described the optimisation of 1,5-diarylpyrrole derivatives toward MTB inhibition.⁵³ In the synthetic route, a Stetter reaction was employed between cuminaldehyde and methyl vinyl ketone (Scheme 7). The reaction was performed under microwave irradiation and driven by the addition of 3-ethyl-5-(2-hydroxyethyl)-4-methylthiazolium bromide which gave the 1,4-diketone 11 in good yield. In the

Scheme 8 Synthesis of MTB inhibitors via a microwave promoted Wittig reaction.

next step, the diketone 11 was cyclised under Paal–Knorr conditions with 4-chloroaniline yielding the tri-substituted pyrrole 12. By the use of a dedicated microwave reactor and septa-sealed reaction vials, the temperature could safely be raised to 160 °C, double the boiling point of the solvent (EtOH), reaching full conversion within 30 min. One of the most active compounds 13 was twice as effective as isoniazid on wild type MTB. The inhibitor 13 was also active towards isoniazid- and rifampicin resistant MTB (MIC = 8.0 and 0.125 μg mL⁻¹ respectively). Recently, the group presented further exploration of the 1,5-diarylpyrrole derivatives.^{52,59}

For further condensation reactions under microwave heating in the synthesis of MTB inhibitors see the following reports by Bairwa et al.,⁶⁰ Torres et al.⁶¹ and Casagnolo et al.⁶²

A Wittig type cyclisation to afford benzofurans was exploited by Alvety et al. in the synthesis of MTB inhibitors (Scheme 8).⁵⁴ It was found that isolation of phosphonium salt increased the yields and furthermore, the reaction times could be shortened from 88 h to 1 h with the use of microwave irradiation compared to traditional reflux. In the one-pot two-step synthesis, the hydroxyl was first coupled with acids or acyl chlorides. In the second step, the esters were then cyclised in a Wittig reaction affording the benzofurans exemplified by 14 (Scheme 8). Compound 14 was also one of the most active compounds which displayed moderate activity in the whole bacterial assay compared to isoniazid.

A structure–activity relationship study of antitubercular nitroimidazoles was carried out by Kim and co-workers in 2009.⁵⁸ In one of the synthetic routes an intramolecular S_N Ar reaction was conducted under single-mode microwave irradiation (Scheme 9). By heating a DMF solution of the amino-functionalised 2-chloroimidazole 15 in the presence of a mild base, at a temperature of 150 °C for 30 min, compound 16 was prepared in modest yield. Inhibitor 16 was evaluated in a whole bacterial assay and displayed a low MIC and furthermore, a related structure in the study was 20 times more potent (MTB, MIC = 0.039μ M).

Proteasomes are involved in the degradation of proteins and have been exploited as targets in a number of human deseases.⁶³ As the proteasomal enzymes are structurally conserved in both eukaryotes and prokaryotes, selectivity is crucial to avoid

Scheme 9 Synthesis of MTB inhibitors via a microwave-assisted intramolecular S_NAr reaction.

Scheme 10 Synthesis of a MTB proteasome inhibitor by a singlemode batch reactor or by a continuous flow non-resonant microwave apparatus.

cytotoxicity. The resolved crystal structure of MTB proteasome⁶⁴ in 2006 improves the prospect for designing MTB specific inhibitors.

In 2009, Lin *et al.* reported the results and exploration of the hits from a HTS targeting the MTB proteasome.⁶⁵ The active compounds comprised an oxathiazol-2-one ring that was found to react covalently but reversibly with the enzyme. In the cyclisation an aryl/heteroaryl-amide was reacted with chlorocarbonylsulfenyl chloride at ambient temperature for 16 h, affording the 5-substituted oxathiazol-2-one in good yield. With the help of a dedicated microwave reactor, the reaction times could be shortened to only 15 min at 100 °C with comparable yields. The 5 phenyl-oxathiazol-2-one 17 was synthesised by the single-mode microwave-assisted method (Scheme 10) and was one of most active compounds towards MTB proteasome displaying a 13 fold selectivity ratio for the rate of inactivation (k_{obs}) over the human proteasome.

Öhrngren and co-workers have evaluated a novel non-resonant microwave reactor for continuous-flow chemistry applications.¹⁸ The non-resonant irradiation heats the entire reactor vessel without pronounced hot or cold spots. The instrument allows high level of safety thanks to a limited reaction zone and fast access to optimised protocols, ready to scale-out. One of the reported applications was the synthesis of the 5-substituted 1,3,4-oxathiazol-2-one 17 (Scheme 10). The reaction was performed with two stock solutions of benzamide and

Scheme 11 Microwave-assisted synthesis of diketo acid HIV-1 integrase inhibitors.

chlorocarbonylsulfenyl chloride that were pumped through a mixer before reaching the microwave applicator. With the optimised conditions; flow rate of 353 μL min⁻¹, residence time 1 min and a temperature of 200 °C, the product 17 was isolated in 62% yield with a throughput of 3.3 mol h^{-1} .

2.2 HIV/AIDS

Human immune deficiency virus (HIV) was identified 30 years ago and since then acquired immune deficiency syndrome (AIDS) has claimed more than 25 million lives. Today, more than 7000 people, including 100 children, are newly infected with the virus each day.⁶⁶ The current treatment consists of highly active antiretroviral therapy (HAART) where patients are given combinations consisting of three or more drugs belonging to two or more classes. These classes include the nucleoside/ nucleotide reverse transcriptase inhibitors, non-nucleoside reverse transcriptase inhibitors, protease inhibitors, fusion inhibitors, CCR5 antagonists and integrase inhibitors.

The integrase enzyme (IN) is used by the virus to insert the pro-viral DNA into the host celluar DNA and thus plays a critical role in the virus life-cycle. Today, raltegravir is the only IN inhibitor approved for clinical use. Chimiri and co-workers have described the synthesis of 1H-benzylindole IN inhibitors where microwave irradiation was employed in three of the four steps in the reaction pathway (Scheme 11).^{67,68} The synthesis commenced with the treatment of commercially available 4-methoxyindole with acetyl chloride in the presence of diethylaluminum chloride. The 3-acylated derivative 18 was obtained after only 10 min of microwave heating at 50 °C compared to 2 h under conventional conditions. Intermediate 18 was then N-alkylated with various benzyl bromides and microwave irradiation was shown to again reduce reaction times (5 min vs. 30 min) and

Scheme 12 Microwave-assisted Leimgruber–Batcho synthesis.

Scheme 13 Microwave-assisted synthesis of HIV IN inhibitor 24 via an ester aminolysis.

also allow the use of less solvent. The 1-benzyl-1H-indoles 19 were then effectively coupled with diethyl oxalate after only 2 min of microwave heating at 50 °C. Finally, the chelating diketo acid group was unmasked under basic conditions to provide the free inhibitor. One of the best compounds 21 showed nanomolar inhibition of the integrease enzyme and HIV-1 replication and had a selectivity index of 228 (Scheme 11).

For additional examples of microwave-assisted S_N2 N-alkylation reactions in the preparation of anti-HIV compounds see Tang et al.⁶⁹ Monforte et al.⁷⁰ and De Luca et al.⁷¹

The Leimgruber–Batcho synthesis^{72,73} is a widely used strategy for the formation of indoles from ortho-nitrotoluenes. This approach was recently utilised in the synthesis of azaindole hyroxamic acid inhibitors of HIV-1 IN (Scheme 12).⁷⁴ The azaindole ring system was assembled by the microwave-assisted reaction of nitropicoline with dimethylformamide dimethyl acetate at 140 °C for 20 min followed by a palladium-catalysed reduction/cyclisation of the enamine intermediate. Subsequent N-alkylation of 22 and installation of the hydroxamic acid moiety gave the final test compounds. The most promising inhibitor 23 exhibited nanomolar enzymatic and anitiviral activity, however the compound series displayed low metabolic stability due to rapid clearance through phase 2 glucuronidation pathways.

In 2009, Yoshinaga and co-workers disclosed a series of 7 benzyl-4-hydroxyisoquinolin-2(1H)-one IN inhibitors where a microwave-assisted ester aminolysis approach was used to introduce various carboxamides into the $C-3$ position.⁷⁵ Compound

Scheme 14 Microwave-assisted triazole formation from a nitrile and an acyl hydrazide.

Scheme 15 The synthesis of S-DABO derivatives using a microwaveassisted Mitsunobu reaction.

24 (Scheme 13) displayed single-digit nanomolar activity in both enzymatic (HIV integrase) and cellular antiviral assays and was also found to possess reasonable bioavailability (20–25%) and acceptable *in vivo* clearance (11.8 mL min⁻¹ kg⁻¹).

A microwave-assisted triazole formation was employed by Johns et al. in the preparation of 26, a potent IN inhibitor and anti-HIV agent (Scheme 14).⁷⁶ Compound 26 was synthesised in moderate yield by the condensation of nitrile 25 with 4-fluorophenylacetic hydrazide at 200 °C.

For additional examples of microwave-assisted heterocyclisations in the synthesis of anti-HIV compounds, see Mehta et al.,⁷⁷ Zhao et al.,⁷⁸ Pace et al.⁷⁹ and Fisher et al.⁸⁰

The reverse transcriptase (RT) of HIV-1 plays an essential role in the synthesis of viral DNA and is the target of sixteen FDA approved drugs. Despite the large number of drugs already on the market, the emergence of resistant virus strains has catalysed the continued search for new RT inhibitors.

In 2009, the Botta group reported the synthesis and anti-HIV activity of a number of chiral cyclopropyl S-DABO (dihydroalkylthio-benzyl-oxopyrimidine) derivatives.⁸¹ The inhibitors were synthesised via a microwave-assisted Mitsunobu reaction between enantiopure cylcopropyl derivatives 27 and 2-thiouracil 28 (Scheme 15). The pure diastereomers were then separated by

Scheme 16 A microwave-assisted three-component synthesis of thiazolidin-4-one derivatives.

Scheme 17 Microwave-assisted asymmetric molybdenum-catalysed alkylation in the synthesis of the HIV-1 protease inhibitor tipranivir.

HPLC and tested for their anti-HIV activity in MT-4 cells. The (R, R, R) isomer 31 was the most potent derivative with picomolar activity against wildtype virus strain and nanomolar activity against three tested mutant strains. The Corelli group has also described the synthesis of S-aryl-S-DABO based HIV-1 inhibitors, synthesised through a microwave-assisted Ullmann type reaction of 2-thiouracils and various arylboronic acids.⁸²

A microwave-assisted three component reaction has been used in the synthesis of 2,3-diaryl-1,3-thiozolidin-4-ones possessing HIV-RT inhibitory activity. 83 The inhibitors were prepared by heating various 2-aminopyridines, benzaldehydes and mercaptoacetic acid in the presence of DCC. The two most potent inhibitors 32 and 33 (Scheme 16) displayed submicromolar inhibitory activity in a HIV-RT kit assay. The same group has also reported the synthesis of dinucleoside substituted

Scheme 18 Microwave-assisted synthesis of tetronic acid analogues of tipranivir.

thiazolidin-4-ones via a microwave-assisted one-pot Staudinger/ aza-Wittig/cyclisation sequence.^{84,85}

Inhibition of the HIV protease enzyme has been a successful strategy in the fight against HIV/AIDS and today there are 10 HIV-1 protease inhibitors on the market. The enzyme is responsible for cleavage of the viral polyprotein into functional enzymes and structural proteins in the final stages of viral replication and is essential for the production of new mature virions.

In work directed towards a stereoselective synthetic route to the HIV-1 protease inhibitor tipranavir, Trost and Andersen used a microwave-assisted molybdenum-catalysed allylic alkylation to prepare one of the key intermediates 34 (Scheme 17). The reaction time was reduced from 24 h at 67 °C to 20 min at 180 °C with only a small reduction in ee (from 96% ee to 94% ee).⁸⁶ In addition, more convenient $Mo(CO)_{6}^{87,88}$ could be used as the molybdenum source instead of the sensitive pre-catalyst complex $Mo(CO)_{3}(C_{7}H_{8}).$

A microwave-assisted S_N2 alkylation/Claisen rearrangement sequence has been utilised by Schobert et al. in the synthesis of tetronic acid analogues of tipranavir.⁸⁹ The most active derivative 37 displayed low micromolar activity against the protease-sensitive virus strain HIV-NL4-3 (Scheme 18).

Hallberg, Larhed and co-workers have, over the past decade, published a number of studies on the microwave-assisted synthesis of HIV-1 protease inhibitors.^{14,90} In general, these have involved the metal-catalysed functionalisation of a target scaffold, with the aid of microwave heating. A range of different scaffolds containing linear and cyclic $1,2$ -dihydoxyethylene $91-99$ and tertiary alcohol^{100–104} transition-state mimicking groups and various palladium-catalysed transformations, including Suzuki– Miyaura, Mizoroki–Heck, Negishi, Stille, Sonogashira, cyanation and aminocarbonylation reactions, have all been explored using this strategy. In a recent example, a microwave-accelerated Stille-coupling was used to prepare 39, a compound possessing potent antiviral activity (Scheme 19, $EC_{50} = 7$ nM).¹⁰³ Compound 39 was synthesised in a moderate yield (40%) by treating

Scheme 19 Microwave-assisted synthesis of HIV-1 protease inhibitor 39 via a Stille cross-coupling.

Scheme 20 Microwave-assisted preparation of an anti-HIV DCP analogue.

aryl bromide 38 with 2-pyridyl tributyltin, $Pd(PPh₃)₂Cl₂$ and CuO at 120 °C for 50 min. Importantly, 39 was only slowly degraded by metabolic enzymes and also retained potency against several resistant HIV-1 isolates.

Microwave-assisted metal-catalysed coupling-reactions have also been utilised by Wiscount *et al.*¹⁰⁵ and Williams *et al.*¹⁰⁶ in the preparation of HIV inhibitors.

A microwave-assisted one-pot alkylation/cyclisation protocol for the synthesis of 2′-2′-dimethyldihydropyranochromone (DCP) containing compounds has recently been developed by the Lee group (Scheme 20).^{107,108} This method was subsequently applied to the synthesis of a library of anti-HIV agents, from which 42 was identified as the most promising drug candidate. Compound 41 was synthesised by treating the resorcinol derivate 40 with 4,4-dimethoxy-2-methyl-2-butanol in the presence of pyridine at 220 °C for 4 h. Further synthetic manipulations led to 42, which showed potent anti-HIV activity against both non-drug-resistant $(HIV-1_{NLA-3})$ and drug-resistant (HIV-RTMDR1) viral strains.

Scheme 21 Synthesis of a CO analogue via a microwave-assisted S_NAr reaction.

2.3 Malaria

According to the WHO, over 3 billion people worldwide live under the threat of malaria infection and in 2009, nearly 800,000 people lost their lives to the disease.^{109,110} Malaria is caused by infection with four species of the family Plasmodium; Plasmodium falciparum (Pf), Plasmodium malariae (Pm), Plasmodium *ovale* (Po) and Plasmodium vivax (Pv) of which Pf is responsible for the most severe and fatal form of the disease.† Most of the drugs currently used in the treatment of malaria target either the haem detoxification system (*e.g.* chloroquine and mefloquine) or the biosynthesis of nucleotide bases by inhibiting dihydrofolate reductase (DHFR) or dihydropteroate reductase (DHPR) (e.g. pyrimethamine/sulfadoxine). The mode of action of artimisenin and its derivitives is still the subject of considerable debate.¹¹¹

Chloroquine (CQ) has been the anti-malaria drug of choice for many decades, however the emergence of CQ-resistance has significantly reduced its efficacy. Thus, the synthesis of CQ analogues with increased activity against CQ-resistant parasite strains has been investigated by a number of research groups. In 2007, Monti and co-workers reported the synthesis of a library of 4-aminoquinolines *via* microwave-assisted S_NAr reactions of commercially available 4,7-dichloroquinoline with various amines (Scheme 21).¹¹² The use of sealed vials and single-mode microwave irradiation was found to decrease reaction times and allowed the reaction to be carried out with aromatic amine nucleophiles. The most promising compound 44 was more active than CQ on a W2 (CQ-resistant) strain of Pf (Scheme 21).

The following year, the same group used microwave-assisted S_N Ar reactions to construct a library of triamino-substituted triazines as analogues of both CQ and cycloguanil (Scheme 22).¹¹³ The target compounds were prepared from cyanuric chloride by three consecutive S_N Ar reactions, the first two conducted at 0 °C and room temperature, respectively, and the final reaction at 180 °C using high density microwave irradiation. The most active compound 45 displayed similar CQ-sensitive (CQ-S) activity and higher CQ-resistant (CQ-R) activity compared to CQ.

Microwave-assisted S_NAr reactions have also been utilised by a number of other groups in the synthesis of antimalarial agents.114–¹¹⁷

Scheme 22 Synthesis of anti malarial triamino-substituted triazines via a microwave heated sequential S_N Ar reactions.

Scheme 23 Synthesis of CQ analogues via a microwave enhanced trimethylaluminium-mediated amide bond formation.

Recently, Sparatore and co-workers described the synthesis and evaluation of CQ and amodiaquine analogues, where a pyrrole ring was introduced into the linker between the 7-chloro-4-aminoquinoline moiety and the basic amine head group.^{118,119} A microwave-assisted trimethylaluminium-mediated amide bond formation was used to install the key basic head group in a number of derivatives, including 47, one of the most active compounds (Scheme 23).¹¹⁹ The use of single-mode microwave heating allowed the reaction time to be reduced from 24 h to 35 min and also improved the yield $($ <10% *vs.* 49%) when compared to classical heating conditions. Compound 47 displayed comparable activity to CQ against the D-10 (CQ-S) strain and better activity than CQ on the W2 (CQ-R) parasite strain of Pf. In addition, a microwave-assisted DCC mediated amide coupling was used to attach the basic head group in a number of less potent compounds.

In 2010, Kabri et al. reported the preparation of 4-anilinoqunazoline derivatives where the majority of the synthetic steps employed microwave heating.¹²⁰ The synthesis of the most active series began with the treatment of 2-aminobenzamide with an excess of chloroacetylchloride (Scheme 24). The intermediate chloroacetamide was then directly cyclised upon reaction with $K₂CO₃$ in water. This two-step microwave-assisted procedure afforded 2-chloromethylquinazolin-4(3H)-one (48) in excellent

Scheme 24 Synthesis of 4-aminoquinazoline 51 *via* a multistep microwave-assisted reaction sequence.

Scheme 25 Synthesis of 4-aminoquinazoline 54 *via* a microwaveassisted coupling-isomerisation reaction.

yield (86% over two steps).¹²¹ Following a standard nitration, treatment with 4-methylbenzenesulfonyl chloride, under microwave conditions, led to the S-alkylated intermediate 49.¹²² Microwave-assisted chlorination and a nucleophilic substitution of the chlorine with various anilines furnished the target library. The most potent member of the series 51 was marginally less active than CQ on the W2 CQ-R parasite strain.

Naphthyridine 54 (Scheme 25), a compound with modest antiplasmodial activity (Pf IC₅₀ = 1.64 mg mL⁻¹), was prepared by a microwave-assisted coupling-isomerisation reaction (MACIR).¹²³ Mechanistically, the reaction is believed to occur via an initial Sonogashira coupling of iodide 52 and propargylic alcohol 53 followed by a base-catalysed propargyl alcohol-enone isomerisation to afford a chalcone intermediate. Finally, a basemediated trans–cis isomerisation and subsequent intramolecular cyclisation generates the naphthyridine ring system.

Very recently, Milner et al. have utilised microwave-assisted nucleophilic epoxide ring opening reactions to prepare a library of diamine quinoline methanol derivatives, based on the mefloquine (MQ) scaffold (Scheme 26).¹²⁴ The most promising candidate 57 was prepared by reacting the enantiopure epoxide 55 with amine 56 under microwave conditions, followed by an acid mediated deprotection of the Boc group. Compound 57 exhibited similar potency to MQ across a range of drug resistant Pf strains and after single dose administration in mice, cured 4 out of

Scheme 26 Synthesis of MQ analogue 57 via a microwave-assisted epoxide ring opening.

Scheme 27 Synthesis of a Pf dUTPase inhibitor via microwaveassisted S_N1 reaction.

5 mice at an orally administrated dose of 320 mg kg−¹ without evidence of toxicity.

For additional examples of microwave promoted nucleophilic ring opening reactions see Robin *et al.* (epoxide opening)¹²⁵ and D'hooghe et al. (aziridine opening).¹²⁶

Gilbert and co-workers have reported the discovery of acyclic uracil-based inhibitors of the Pf enzyme deoxyuridine nucleotidohydrolyase $(dUTPase)^{127}$ A microwave-assisted nucleophilic substitution was used to produce the tritylated compound 58 (Scheme 27) and interestingly, attempts to conduct the reaction under classical heating conditions failed. Compound 58 was found to selectively inhibit Pf dUTPase ($K_i = 2.5 \mu M$ vs. human dUTPase $K_i > 1000 \mu M$) and inhibit the growth of Pf in vitro $(IC_{50} = 0.9 \mu M).$

As part of a screening campaign aimed at identifying smallmolecules with whole-cell antimalarial activity, Mazitshek and co-workers discovered a 2-amino-3-hydroxyindole derivative with good in vivo efficacy (Scheme 28).¹²⁸ To facilitate rapid analogue generation, the authors sought to develop a short and robust synthetic method for the preparation of the unusual core structure. The target compounds were accessed in only two steps starting with a Grignard addition to commercially available isatins. The oxindole intermediate was then treated with tertbutyldimethylsilyl amine (TBDMSNH₂) and SnCl₄ under microwave irradiation, in a one-pot amidine formation/deprotection reaction, to afford the desired 2-amino-3-hydroxyindoles in

Scheme 28 Microwave-assisted synthesis of a anti malarial 2-amino-3 hydroxyindole derivative.

Scheme 29 Microwave-assisted cross metathesis in the synthesis of a Falcipain-2 inhibitor.

reasonable yields. Notably, this was the first reported use of $TBDMSNH₂$ as an ammonia equivalent and no loss of enantiomeric excess was detected when the reaction was carried out with an enantioenriched oxindole. The most active compound 59 (Scheme 28) was more active than CQ on a drug-resistant parasite strain and slightly less potent than CQ on a drug-sensitive strain.

Falcipain-2 (FP-2) is a cysteine protease involved in haemoglobin degradation, a process that provides essential nutrients for parasite growth¹²⁹ and cleavage of host cytoskeleton proteins, leading to cell rupture and merozioite release.¹³⁰ The Zappalà group have utilised microwave-assisted cross metathesis reactions as a key step in the synthesis of peptidomimetic inhibitors of FP-2.131–¹³³ The cross-metathesis reaction was used to introduce various α,β-unsaturated electrophilic groups onto a 1,4 benzodiazepine scaffold and the synthesis of one of the most potent compounds 60 (FP-2 $K_i = 17$ nM and $PfIC_{50} = 12 \mu M$) is depicted in Scheme 29.

The plasmepsins are a group of 10 aspartic acid proteases encoded in the Pf genome and four of these, plasmepsin I (Plm1), plasmepsin II (Plm2), plasmepsin IV (Plm4) and histoaspartic protease (HAP or Plm3), are involved in haemoglobin degradation. In 2003, Hallberg and co-workers presented the $first^{134}$ of a series papers describing the single-mode microwave-

Scheme 30 Synthesis of a plasmepsin I and II inhibitor via a microwave induced Suzuki–Miyaura reaction.

assisted synthesis of plasmepsin inhibitors.^{13,135–137} In this work, a hydroxyethylamine transition-state isostere was utilised and different P1 side chains were evaluated via the decoration of the benzylic side chains (Scheme 30). Inhibitors containing either a para-bromo or a meta-triflate group in the P1-position served as starting materials for microwave-assisted Suzuki– Miyaura reactions with four different arylboronic acids. In total, eight inhibitors were produced after 20 min of irradiation, however the isolated yields were only moderate. The most potent inhibitor of the series (61) displayed a K_i value of 63 nM for Plm I and 150 nM for Plm II, with moderate selectivity versus Cat D (Scheme 30).

For additional examples of microwave-asssisted metal-catalysed coupling reactions in the synthesis of antimalarial agents see Choi et al. (cyanation)¹³⁸ Bulbule et al. (cyanation)¹³⁹ Bouloc et al. (Suzuki-Miyaura)¹⁴⁰

In 2009, Orrling et al. described the synthesis of Plm4 inhibitors containing an α-substituted norstatine transition-state isostere.¹⁴¹ The inhibitors were prepared by a convergent sequence where two key intermediates (64 and 66) were assembled with the aid of microwave heating (Scheme 31). The 5,5 dimethylthiazolidine moiety (62 and 63) was generated by treating penicillamine with formaldehyde at 110 °C, which resulted in a reduction of the reaction time from 16 h to 5 min when compared to the previously reported room temperature procedure.¹⁴² The α -substituted norstatine core 66 was derived from a substituted acrylic acid ester 65, prepared in a one-pot esterification of the corresponding acid using $S OCl₂$ and ethanol. The best compound in the series (67) displayed nanomolar K_i values against Plm4 from all four disease causing parasite species.

2.4 Hepatitis C

In 2009, it was estimated that 130–170 million people or 2.2–3% of the world's population were infected with hepatitis C (HCV).143 One in every forty deaths is related to HCV and the virus accounts for 27% of all cirrhosis deaths and 25% of all hepatocellular carcinoma deaths worldwide.¹⁴⁴ The current

Scheme 31 Microwave-assisted synthesis of a plasmepsin IV inhibitor.

HCV-treatment is a combination therapy with pegylated interferon alfa and ribavirin together with one of the newly FDA approved drugs, boceprevir or telaprevir, which also are the first HCV specific drugs that have reached the market. Boceprevir and telaprevir target the HCV NS3 protease which is involved in viral polyprotein processing that releases the essential structural proteins and viral enzymes required for viral replication.¹⁴⁵

Nilsson and co-workers explored a series of HCV NS3 inhibitors that were cyclised between the P1 and P3 residue (Scheme 32).¹⁴⁶ A microwave-assisted ring opening of the tricyclic lactone followed by amide coupling gave the key intermediate 68 in good yield over three steps with retained stereochemistry. After O-arylation, the ethyl ester was hydrolysed under basic conditions using microwave heating. A twostep microwave-assisted protocol was then applied to couple the resulting acid with cyclopropanesulfonamide followed by macrocyclic ring closing metathesis to yield 70. The macrocyclic inhibitor 70 showed subnanomolar affinity to the HCV NS3 protease.

There are additional examples of NS3 protease inhibitors synthesised via microwave accelerated metathesis macrocyclisations between $P2-P1'$ ¹⁴⁷ and between $P2-P4'$ ¹⁴⁸ in the literature.

In 2009, Gising et al. developed a microwave-assisted one-pot two-step protocol to afford N-1 and C-6 functionalised 3,5 dichloro- $2(1H)$ -pyrazinones with an application towards HCV NS3 protease inhibition (Scheme 33). 149 The products were isolated in comparable yields with the literature and the reaction time was shortened to 2 times 10 min instead of several hours with traditional heating. In the one-pot two-step sequence an amine, an aldehyde and trimethylsilyl cyanide were cyclised together with oxalyl chloride to afford a set of 3,5-dichloro-2

Scheme 32 Microwave-assisted synthesis of HCV NS3 protease inhibitors via metathesis.

74; HCV NS3 protease $K_i = 6.8 \pm 1.1 \,\mu M$

Scheme 33 Synthesis of HCV NS3 protease inhibitors via microwave mediated multi-component heterocyclisation and S_NAr substitution.

 (1) -pyrazinones exemplified by 71 in Scheme 33.¹⁴⁹ The following year Örtqvist and co-workers reported a number of 2 $(1H)$ -pyrazinone based HCV NS3 protease inhibitors.¹⁵⁰ Singlemode microwave heating was utilised to selectively substitute the C-3 chlorine with 3,3-dimethylbutanamine yielding 72 and hydrolysis under controlled microwave irradiation gave 73 quantitatively. The final inhibitor 74 was reached after peptide coupling and showed moderate affinity to the NS3 protease.

In 2005, Wu et al. explored palladium-catalysed amidocarbonylation reactions using septa-sealed microwave vials and $Mo(CO)₆$ as a solid source of carbon monoxide. The developed protocol was also employed in the synthesis of a NS3 protease inhibitor 76 (Scheme 34).¹⁵¹ The scope of the reaction was further investigated by Rönn and co-workers in 2008, exploiting more complex aryl bromides as NS3 protease inhibitor precursors in 11 related examples.¹⁵²

Another viral target under investigation is the NS5B polymerase that catalyzes RNA synthesis during replication. NS5B is a validated target and is crucial for viral infectivity.¹⁵³

A recent article from Ontoria et al. described the results from a HTS targeting the NS5B polymerase and the further exploration of the resulting hits.¹⁵⁴ The re-synthesis and exploration of the hits started with a condensation between the cyclic anhydride 77 and 3-bromoaniline. The reaction required heating in NMP for 24 h at 145 °C to reach full conversion giving the product in moderate yield. The use of microwave heating of sealed reactions increased the yield and decreased the reaction time to 30 min at 200 °C. A subsequent S_NAr substitution of 78 with various amines produced a set of NS5B polymerase inhibitors and the most promising compound 79 (Scheme 35) was more potent towards HCV genotype 1 than to genotype 2.

Recently, Lazerwith co-workers described the pharmacokinetic optimisation of a HCV replicon inhibitor.¹⁵⁵ One of the most promising compounds was synthesised through a

Scheme 34 Microwave-assisted synthesis of HCV NS3 protease inhibitors via a palladium-catalysed amidocarbonylation.

Scheme 35 Synthesis of HCV NS5B polymerase inhibitors via a microwave promoted condensation reaction.

palladium-catalysed, microwave mediated Suzuki–Miyaura cross-coupling¹⁵⁶ (Scheme 36). Compound 80 displayed improved bioavailability ($F = 88\%$ vs. 2%) and prolonged mean residence time (MRT = 3.5 h vs. 0.4 h) in rat.

Other successful microwave-assisted metal-catalysed crosscouplings for the synthesis of HCV inhibitors have been reported by several groups and include; Suzuki–Miyaura cross-couplings,^{157,158} Sonogashira couplings,^{159,160} Negishi cross-coupling¹⁶¹ and Cu-mediated cyanation.¹⁶²

Arbidol (see Scheme 37) is a broad spectrum antibiotic and a known HCV entry inhibitor.¹⁶³ In a report from 2010, Selitto and

Scheme 36 Synthesis of HCV replicon inhibitors via a microwave heated Suzuki–Miyaura cross-coupling.

Huh-7.5 cells TC_{50} > 50 µM arbidol; HCV entry inhibition, genotype 2a, $IC_{50} = 5 \mu M$ Huh-7.5 cells $TC_{50} = 12 \mu M$

Scheme 37 Synthesis of HCV entry inhibitors via a microwave promoted reductive amination.

co-workers described the synthesis and evaluation of arbidol analogues.¹⁶⁴ In the final synthetic step, a set of amines was introduced by reductive amination in sealed vials under controlled microwave irradiation (Scheme 37). One of the most active compounds 81, showed comparable anti HCV entry inhibition to arbidol and a better cytotoxicity profile $(TC_{50} > 50 \mu M)$ vs. arbidol, $TC_{50} = 12 \mu M$).

3. Conclusions

Controlled and automated single-mode microwave heating is one constituent of the collection of novel techniques that provide medicinal chemists with new and powerful production tools. In this perspective we have summarised a number of diverse microwave-assisted synthetic protocols that have been useful in discovery projects towards the development of new antibiotics and antiviral compounds. In many of the depicted examples, the main chemical effort was directed towards fast, smooth and reliable small-scale procedures avoiding complicated handling of inert conditions and reactive gases. Furthermore, the microwave methodology helped establish the structure–activity relationships in a uniquely rapid fashion. Thus, the field of organic synthesis is evolving and controlled microwave heating will continue to change the way we perform both target and diversity driven medicinal chemistry. Today, microwave chemistry accelerates and increases the opportunities to develop not only new antibiotics and antiviral drugs but all kinds of new medicines. 3 **Concellusion S**

Concellular at the control of the control of the control of the control of New York at Albany on 24 March 2012 Published on 24 March 2012 Published on 24 March 2012 Published on 24 March 2012 Published

Acknowledgements

We would like to express our sincere gratitude to the Swedish Research Council and to Knut and Alice Wallenberg's Foundation.

Notes and references

- †All biological data refers to Pf unless otherwise stated.
- 1 R. J. Giguere, T. L. Bray, S. M. Duncan and G. Majetich, Tetrahedron Lett., 1986, 27, 4945–4948.
- 2 R. Gedye, F. Smith, K. Westaway, H. Ali, L. Baldisera, L. Laberge and J. Rousell, Tetrahedron Lett., 1986, 27, 279–282.
- 3 C. Strauss and R. Trainor, Aust. J. Chem., 1995, 48, 1665–1692.
- 4 P. Lidstrom, J. Tierney, B. Wathey and J. Westman, Tetrahedron, 2001, 57, 9225–9283.
- 5 C. O. Kappe, Angew. Chem., Int. Ed., 2004, 43, 6250–6284.
- 6 P. Nilsson, K. Olofsson and M. Larhed, Top. Curr. Chem., 266, 103–144.
- 7 S. Caddick and R. Fitzmaurice, Tetrahedron, 2009, 65, 3325–3355. 8 C. O. Kappe and A. Stadler, Microwaves in Organic and Medicinal Chemistry, Wiley-VCH, Weinheim, 2005.
- 9 C. O. Kappe and D. Dallinger, Mol. Diversity, 2009, 13, 71–193.
- 10 N. Elander, J. R. Jones, S. Y. Lu and S. Stone-Elander, Chem. Soc. Rev., 2000, 29, 239–249.
- 11 M. A. Herrero, J. M. Kremsner and C. O. Kappe, J. Org. Chem., 2008, 73, 36–47.
- 12 A. Stadler, B. H. Yousefi, D. Dallinger, P. Walla, E. Van der Eycken, N. Kaval and C. O. Kappe, Org. Process Res. Dev., 2003, 7, 707–716.
- 13 D. Nöteberg, W. Schaal, E. Hamelink, L. Vrang and M. Larhed, J. Comb. Chem., 2003, 5, 456–464.
- 14 M. Larhed and A. Hallberg, Drug Discovery Today, 2001, 6, 406–416.
- 15 I. R. Baxendale, C. M. Griffiths-Jones, S. V. Ley and G. K. Tranmer, Chem.–Eur. J., 2006, 12, 4407–4416.
- 16 E. Comer and M. G. Organ, J. Am. Chem. Soc., 2005, 127, 8160–8167.
- 17 J. D. Moseley and E. K. Woodman, Org. Process Res. Dev., 2008, 12, 967–981.
- 18 P. Öhrngren, A. Fardost, F. Russo, J. S. Schanche, M. Fagrell and M. Larhed, Unpublished work
- 19 J. R. Schmink, C. M. Kormos, W. G. Devine and N. E. Leadbeater, Org. Process Res. Dev., 2010, 14, 205–214.
- 20 D. Dallinger, H. Lehmann, J. D. Moseley, A. Stadler and C. O. Kappe, Org. Process Res. Dev., 2011, 15, 841–854.
- 21 The Global Burden of Disease: 2004 Update, World Health Organization, Geneva, 2008
- 22 Global Tuberculosis Control 2010, World Health Organization, Geneva, 2010
- 23 G. B. Migliori, C. Lange, R. Centis, G. Sotgiu, R. Mutterlein, H. Hoffmann, K. Kliiman, G. De laco, F. N. Lauria, M. D. Richardson, A. Spanevello, D. M. Cirillo and T. S. Grp, Eur. Respir. J., 2008, 31, 1155–1159.
- 24 M. C. Raviglione and I. M. Smith, N. Engl. J. Med., 2007, 356, 656–659.
- 25 G. Harth, D. L. Clemens and M. A. Horwitz, Proc. Natl. Acad. Sci. U. S. A., 1994, 91, 9342-9346.
- 26 D. L. Clemens and M. A. Horwitz, J. Exp. Med., 1995, 181, 257–270.
- 27 A. H. Gordon, P. Darcyhart and M. R. Young, Nature, 1980, 286, 79–80.
- 28 G. Harth and M. A. Horwitz, J. Exp. Med., 1999, 189, 1425–1435.
- 29 G. Harth and M. A. Horwitz, Infect. Immun., 2003, 71, 456–464.
- 30 J. Gising, M. T. Nilsson, L. R. Odell, S. Yahiaoui, H. Iyer, M. Lindh, B. R. Srinivasa, M. Larhed, S. L. Mowbray and A. Karlén, J. Med. Chem., Submitted
- 31 L. R. Odell, M. T. Nilsson, J. Gising, O. Lagerlund, D. Muthas, A. Nordqvist, A. Karlen and M. Larhed, Bioorg. Med. Chem. Lett., 2009, 19, 4790–4793.
- 32 T. H. Manjashetty, P. Yogeeswari and D. Sriram, Bioorg. Med. Chem. Lett., 2011, 21, 2125-2128.
- 33 J. C. Sacchettini and C. D. Poulter, Science, 1997, 277, 1788–1789.
- 34 P. J. Proteau, Bioorg. Chem., 2004, 32, 483–493.
- 35 T. Kuzuyama, T. Shimizu, S. Takahashi and H. Seto, Tetrahedron Lett., 1998, 39, 7913–7916.
- 36 S. Takahashi, T. Kuzuyama, H. Watanabe and H. Seto, Proc. Natl. Acad. Sci. U. S. A., 1998, 95, 9879–9884.
- 37 M. Rodriguez-Concepcion, N. Campos, L. M. Lois, C. Maldonado, J. F. Hoeffler, C. Grosdemange-Billiard, M. Rohmer and A. Boronat, FEBS Lett., 2000, 473, 328–332.
- 38 M. Andaloussi, L. M. Henriksson, A. Wieckowska, M. Lindh, C. Björkelid, A. M. Larsson, S. Suresh, H. Iyer, B. R. Srinivasa, T. Bergfors, T. Unge, S. L. Mowbray, M. Larhed, T. A. Jones and A. Karlén, J. Med. Chem., 2011, 54, 4964–4976.
- 39 J. Lindh, P. A. Enquist, A. Pilotti, P. Nilsson and M. Larhed, J. Org. Chem., 2007, 72, 7957–7962.
- 40 A. Nordqvist, C. Björkelid, M. Andaloussi, A. M. Jansson, S. L. Mowbray, A. Karlén and M. Larhed, J. Org. Chem., 2011, 76, 8986–8998.
- 41 M. Andaloussi, M. Lindh, C. Björkelid, S. Suresh, A. Wieckowska, H. Iyer, A. Karlén and M. Larhed, Bioorg. Med. Chem. Lett., 2011, 21, 5403–5407.
- 42 R. Singh, V. Rao, H. Shakila, R. Gupta, A. Khera, N. Dhar, A. Singh, A. Koul, Y. Singh, M. Naseema, P. R. Narayanan, C. N. Paramasivan, V. D. Ramanathan and A. K. Tyagi, Mol. Microbiol., 2003, 50, 751–762.
- 43 E. H. Fischer, H. Charbonneau and N. K. Tonks, Science, 1991, 253, 401–406.
- 44 L. P. Tan, H. Wu, P. Y. Yang, K. A. Kalesh, X. H. Zhang, M. Y. Hu, R. Srinivasan and S. Q. Yao, Org. Lett., 2009, 11, 5102–5105.
- 45 C. W. Tornøe, C. Christensen and M. Meldal, J. Org. Chem., 2002, 67, 3057–3064.
- 46 V. V. Rostovtsev, L. G. Green, V. V. Fokin and K. B. Sharpless, Angew. Chem., Int. Ed., 2002, 41, 2596–2599.
- 47 P. Kim, L. Zhang, U. H. Manjunatha, R. Singh, S. Patel, J. Jiricek, T. H. Keller, H. I. Boshoff, C. E. Barry and C. S. Dowd, J. Med. Chem., 2009, 52, 1317–1328.
- 48 O. Lagerlund, L. R. Odell, S. L. Mowbray, M. T. Nilsson, W. W. Krajewski, A. Nordqvist, A. Karlen and M. Larhed, Comb. Chem. High. T. Scr., 2007, 10, 783–789.
- 49 K. Manna and Y. K. Agrawal, Eur. J. Med. Chem., 2010, 45, 3831–3839.
- 50 S. V. Karthikeyan, S. Perumal, K. A. Shetty, P. Yogeeswari and D. Sriram, Bioorg. Med. Chem. Lett., 2009, 19, 3006–3009.
- 51 D. Castagnolo, M. Radi, F. Dessi, F. Manetti, M. Saddi, R. Meleddu, A. De Logu and M. Botta, Bioorg. Med. Chem. Lett., 2009, 19, 2203– 2205.
- 52 M. Biava, G. C. Porretta, G. Poce, A. De Logu, R. Meleddu, E. De Rossi, F. Manetti and M. Botta, Eur. J. Med. Chem., 2009, 44, 4734– 4738.
- 53 M. Biava, G. C. Porretta, G. Poce, A. De Logu, M. Saddi, R. Meleddu, F. Manetti, E. De Rossi and M. Botto, J. Med. Chem., 2008, 51, 3644– 3648.
- 54 L. Alvey, S. Prado, B. Saint-Joanis, S. Michel, M. Koch, S. I. Cole, F. Tillequin and Y. L. Janin, Eur. J. Med. Chem., 2009, 44, 2497–2505.
- 55 M. D. Kakwani, N. H. Palsule Desai, A. C. Lele, M. Ray, M. G. R. Rajan and M. S. Degani, Bioorg. Med. Chem. Lett., 2011, 21, 6523–6526.
- 56 Y. Zhou, A. B. Beeler, S. Y. Cho, Y. H. Wang, S. G. Franzblau and J. K. Snyder, J. Comb. Chem., 2008, 10, 534–540.
- 57 K. Balamurugan, V. Jeyachandran, S. Perumal, T. H. Manjashetty, P. Yogeeswari and D. Sriram, Eur. J. Med. Chem., 2010, 45, 682–688.
- 58 P. Kim, S. Kang, H. I. Boshoff, J. Jiricek, M. Collins, R. Singh, U. H. Manjunatha, P. Niyomrattanakit, L. Zhang, M. Goodwin, T. Dick, T. H. Keller, C. S. Dowd and C. E. Barry, J. Med. Chem., 2009, 52, 1329–1344. The Resolution of New York at Albany on 24 March 2012 at Albany of New York at Albany on α March 2012 at Albany on α March 2013
	- 59 M. Biava, G. C. Porretta, G. Poce, C. Battilocchio, S. Alfonso, A. de Logu, F. Manetti and M. Botta, ChemMedChem, 2011, 6, 593–599.
	- 60 R. Bairwa, M. Kakwani, N. R. Tawari, J. Lalchandani, M. K. Ray, M. G. R. Rajan and M. S. Degani, Bioorg. Med. Chem. Lett., 2010, 20, 1623–1625.
	- 61 E. Torres, E. Moreno, S. Ancizu, C. Barea, S. Galiano, I. Aldana, A. Monge and S. Perez-Silanes, Bioorg. Med. Chem. Lett., 2011, 21, 3699–3703.
	- 62 D. Castagnolo, F. Manetti, M. Radi, B. Bechi, M. Pagano, A. De Logu, R. Meleddu, M. Saddi and M. Botta, Bioorg. Med. Chem., 2009, 17, 5716–5721.
	- 63 G. de Bettignies and O. Coux, Biochimie, 2010, 92, 1530–1545.
	- 64 G. Q. Hu, G. Lin, M. Wang, L. Dick, R. M. Xu, C. Nathan and H. L. Li, Mol. Microbiol., 2006, 59, 1417–1428.
	- 65 G. Lin, D. Y. Li, L. P. S. de Carvalho, H. T. Deng, H. Tao, G. Vogt, K. Y. Wu, J. Schneider, T. Chidawanyika, J. D. Warren, H. L. Li and C. Nathan, Nature, 2009, 461, 621–U663.
	- 66 AIDS epidemic update 2009, UNAIDS/09.36E/JC1700E, World Health Organization, Geneva, 2009
	- 67 L. De Luca, M. L. Barreca, S. Ferro, N. Iraci, M. Michiels, F. Christ, Z. Debyser, M. Witvrouw and A. Chimirri, Bioorg. Med. Chem. Lett., 2008, 18, 2891–2895.
	- 68 S. Ferro, L. De Luca, M. L. Barreca, N. Iraci, S. De Grazia, F. Christ, M. Witvrouw, Z. Debyser and A. Chimirri, J. Med. Chem., 2009, 52, 569–573.
	- 69 J. Tang, K. Maddali, M. Metifiot, Y. Y. Sham, R. Vince, Y. Pommier and Z. Wang, J. Med. Chem., 2011, 54, 2282–2292.
	- 70 A.-M. Monforte, A. Rao, P. Logoteta, S. Ferro, L. De Luca, M. L. Barreca, N. Iraci, G. Maga, E. De Clercq, C. Pannecouque and A. Chimirri, Bioorg. Med. Chem., 2008, 16, 7429–7435.
	- 71 L. De Luca, S. Ferro, R. Gitto, M. L. Barreca, S. Agnello, F. Christ, Z. Debyser and A. Chimirri, Bioorg. Med. Chem., 2010, 18, 7515–7521.
	- 72 R. D. Clark and D. B. Repke, Heterocycles, 1984, 22, 195–221.
	- 73 A. D. Bachto and W. Leimgruber, Org. Synth., 1985, 63, 214.
	- 74 M. B. Plewe, S. L. Butler, K. R. Dress, Q. Hu, T. W. Johnson, J. E. Kuehler, A. Kuki, H. Lam, W. Liu, D. Nowlin, Q. Peng, S. V. Rahavendran, S. P. Tanis, K. T. Tran, H. Wang, A. Yang and J. Zhang, J. Med. Chem., 2009, 52, 7211–7219.
	- 75 E. E. Boros, C. E. Edwards, S. A. Foster, M. Fuji, T. Fujiwara, E. P. Garvey, P. L. Golden, R. J. Hazen, J. L. Jeffrey, B. A. Johns, T. Kawasuji, R. Kiyama, C. S. Koble, N. Kurose, W. H. Miller, A. L. Mote, H. Murai, A. Sato, J. B. Thompson, M. C. Woodward and T. Yoshinaga, J. Med. Chem., 2009, 52, 2754–2761.
	- 76 B. A. Johns, J. G. Weatherhead, S. H. Allen, J. B. Thompson, E. P. Garvey, S. A. Foster, J. L. Jeffrey and W. H. Miller, Bioorg. Med. Chem. Lett., 2009, 19, 1802–1806.
	- 77 V. P. Mehta, S. G. Modha, E. Ruijter, K. Van Hecke, L. Van Meervelt, C. Pannecouque, J. Balzarini, R. V. A. Orru and E. Van der Eycken, J. Org. Chem., 2011, 76, 2828–2839.
	- 78 Z. Zhao, S. E. Wolkenberg, M. Lu, V. Munshi, G. Moyer, M. Feng, A. V. Carella, L. T. Ecto, L. J. Gabryelski, M.-T. Lai, S. G. Prasad, Y. Yan, G. B. McGaughey, M. D. Miller, C. W. Lindsley, G. D. Hartman, J. P. Vacca and T. M. Williams, Bioorg. Med. Chem. Lett., 2008, 18, 554– 559.
	- 79 P. Pace, S. A. H. Spieser and V. Summa, Bioorg. Med. Chem. Lett., 2008, 18, 3865–3869.
	- 80 T. E. Fisher, B. Kim, D. D. Staas, T. A. Lyle, S. D. Young, J. P. Vacca, M. M. Zrada, D. J. Hazuda, P. J. Felock, W. A. Schleif, L. J. Gabryelski, M. R. Anari, C. J. Kochansky and J. S. Wai, Bioorg. Med. Chem. Lett., 2007, 17, 6511–6515.
	- 81 M. Radi, G. Maga, M. Alongi, L. Angeli, A. Samuele, S. Zanoli, L. Bellucci, A. Tafi, G. Casaluce, G. Giorgi, M. Armand-Ugon, E. Gonzalez, J. A. Esté, M. Baltzinger, G. Bec, P. Dumas, E. Ennifar and M. Botta, J. Med. Chem., 2009, 52, 840–851.
	- 82 C. Mugnaini, F. Manetti, J. A. Esté, I. Clotet-Codina, G. Maga, R. Cancio, M. Botta and F. Corelli, Bioorg. Med. Chem. Lett., 2006, 16, 3541–3544.
	- 83 H. Chen, J. Bai, L. Jiao, Z. Guo, Q. Yin and X. Li, Bioorg. Med. Chem., 2009, 17, 3980–3986.
- 84 H. Chen, J. Zhao, Y. Li, F. Shen, X. Li, Q. Yin, Z. Qin, X. Yan, Y. Wang, P. Zhang and J. Zhang, Bioorg. Med. Chem. Lett., 2011, 21, 574–576.
- 85 F. Shen, X. Li, X. Zhang, Q. Yin, Z. Qin, H. Chen, J. Zhang and Z. Ma, Org. Biomol. Chem., 2011, 9, 5766–5772.
- 86 B. M. Trost and N. G. Andersen, J. Am. Chem. Soc., 2002, 124, 14320– 14321.
- 87 O. Belda, N. F. Kaiser, U. Bremberg, M. Larhed, A. Hallberg and C. Moberg, J. Org. Chem., 2000, 65, 5868–5870.
- 88 N. F. K. Kaiser, U. Bremberg, M. Larhed, C. Moberg and A. Hallberg, Angew. Chem., Int. Ed., 2000, 39, 3595–3598.
- 89 R. Schobert, R. Stehle and H. Walter, Tetrahedron, 2008, 64, 9401–9407.
- 90 J. Wannberg, K. Ersmark and M. Larhed, Top. Curr. Chem., 266, 167– 198.
- 91 M. Alterman, H. O. Andersson, N. Garg, G. Ahlsén, S. Lövgren, B. Classon, U. H. Danielson, I. Kvarnström, L. Vrang, T. Unge, B. Samuelsson and A. Hallberg, J. Med. Chem., 1999, 42, 3835–3844.
- 92 M. Alterman and A. Hallberg, J. Org. Chem., 2000, 65, 7984–7989.
93 W. Schaal, A. Karlsson, G. Ahlsén, J. Lindberg, H. O. Ander
- 93 W. Schaal, A. Karlsson, G. Ahlsén, J. Lindberg, H. O. Andersson, U. H. Danielson, B. Classon, T. Unge, B. Samuelsson, J. Hultén, A. Hallberg and A. Karlén, J. Med. Chem., 2001, 44, 155–169.
- 94 J. Wannberg, D. Dallinger, C. O. Kappe and M. Larhed, J. Comb. Chem., 2005, 7, 574–583.
- 95 A. Ax, W. Schaal, L. Vrang, B. Samuelsson, A. Hallberg and A. Karlén, Bioorg. Med. Chem., 2005, 13, 755–764.
- 96 J. Wannberg, Y. A. Sabnis, L. Vrang, B. Samuelsson, A. Karlén, A. Hallberg and M. Larhed, Bioorg. Med. Chem., 2006, 14, 5303–5315.
- 97 X. Wu, J. Wannberg and M. Larhed, Tetrahedron, 2006, 62, 4665–4670.
- 98 H. Gold, A. Ax, L. Vrang, B. Samuelsson, A. Karlén, A. Hallberg and M. Larhed, Tetrahedron, 2006, 62, 4671–4675.
- 99 A. Ax, A. A. Joshi, K. M. Orrling, L. Vrang, B. Samuelsson, A. Hallberg, A. Karlén and M. Larhed, Tetrahedron, 2010, 66, 4049– 4056.
- 100 J. K. Ekegren, N. Ginman, Å. Johansson, H. Wallberg, M. Larhed, B. Samuelsson, T. Unge and A. Hallberg, J. Med. Chem., 2006, 49, 1828–1832.
- 101 X. Wu, J. K. Ekegren and M. Larhed, Organometallics, 2006, 25, 1434–1439.
- 102 X. Wu, P. Öhrngren, J. K. Ekegren, J. Unge, T. Unge, H. Wallberg, B. Samuelsson, A. Hallberg and M. Larhed, J. Med. Chem., 2008, 51, 1053–1057.
- 103 A. K. Mahalingam, L. Axelsson, J. K. Ekegren, J. Wannberg, J. Kihlstrom, T. Unge, H. Wallberg, B. Samuelsson, M. Larhed and A. Hallberg, J. Med. Chem., 2010, 53, 607–615.
- 104 P. Öhrngren, X. Wu, M. Persson, J. K. Ekegren, H. Wallberg, L. Vrang, A. Rosenquist, B. Samuelsson, T. Unge and M. Larhed, Med. Chem. Commun., 2011, 2, 701–709.
- 105 C. M. Wiscount, P. D. Williams, L. O. Tran, M. W. Embrey, T. E. Fisher, V. Sherman, C. F. Homnick, D. Donnette Staas, T. A. Lyle, J. S. Wai, J. P. Vacca, Z. Wang, P. J. Felock, K. A. Stillmock, M. V. Witmer, M. D. Miller, D. J. Hazuda, A. M. Day, L. J. Gabryelski, L. T. Ecto, W. A. Schleif, D. J. DiStefano, C. J. Kochansky and M. Reza Anari, Bioorg. Med. Chem. Lett., 2008, 18, 4581–4583.
- 106 P. D. Williams, D. D. Staas, S. Venkatraman, H. M. Loughran, R. D. Ruzek, T. M. Booth, T. A. Lyle, J. S. Wai, J. P. Vacca, B. P. Feuston, L. T. Ecto, J. A. Flynn, D. J. DiStefano, D. J. Hazuda, C. M. Bahnck, A. L. Himmelberger, G. Dornadula, R. C. Hrin, K. A. Stillmock, M. V. Witmer, M. D. Miller and J. A. Grobler, Bioorg. Med. Chem. Lett., 2010, 20, 6754–6757.
- 107 T. Zhou, Q. Shi, C.-H. Chen, H. Zhu, L. Huang, P. Ho and K.-H. Lee, Bioorg. Med. Chem., 2010, 18, 6678–6689.
- 108 T. Zhou, Q. Shi and K. H. Lee, Tetrahedron Lett., 2010, 51, 4382–4386.
- 109 World Malaria Report 2005, World Health Organization, Geneva, 2005
- 110 World Malaria Report 2010, World Health Organization, Geneva, 2010
- 111 P. M. O'Neill, V. E. Barton and S. A. Ward, Molecules, 2010, 15, 1705– 1721.
- 112 S. Melato, P. Coghi, N. Basilico, D. Prosperi and D. Monti, Eur. J. Org. Chem., 2007, 2007, 6118–6123.
- 113 S. Melato, D. Prosperi, P. Coghi, N. Basilico and D. Monti, ChemMed-Chem, 2008, 3, 873–876.
- 114 A. Kumar, K. Srivastava, S. Raja Kumar, S. K. Puri and P. M. S. Chauhan, Bioorg. Med. Chem. Lett., 2010, 20, 7059–7063.
- 115 S. Gemma, G. Kukreja, C. Fattorusso, M. Persico, M. P. Romano, M. Altarelli, L. Savini, G. Campiani, E. Fattorusso, N. Basilico, D. Taramelli, V. Yardley and S. Butini, Bioorg. Med. Chem. Lett., 2006, 16, 5384–5388.
- 116 D. C. Martyn, A. Nijjar, C. A. Celatka, R. Mazitschek, J. F. Cortese, E. Tyndall, H. Liu, M. M. Fitzgerald, T. J. O'Shea, S. Danthi and J. Clardy, Bioorg. Med. Chem. Lett., 2010, 20, 228–231.
- 117 A. J. Ndakala, R. K. Gessner, P. W. Gitari, N. October, K. L. White, A. Hudson, F. Fakorede, D. M. Shackleford, M. Kaiser, C. Yeates, S. A. Charman and K. Chibale, J. Med. Chem., 2011, 54, 4581–4589.
- 118 M. Casagrande, N. Basilico, S. Parapini, S. Romeo, D. Taramelli and A. Sparatore, Bioorg. Med. Chem., 2008, 16, 6813–6823.
- 119 M. Casagrande, N. Basilico, C. Rusconi, D. Taramelli and A. Sparatore, Bioorg. Med. Chem., 2010, 18, 6625–6633.
- 120 Y. Kabri, N. Azas, A. Dumètre, S. Hutter, M. Laget, P. Verhaeghe, A. Gellis and P. Vanelle, Eur. J. Med. Chem., 2010, 45, 616–622.
- 121 Y. Kabri, A. Gellis and P. Vanelle, Green Chem., 2009, 11, 201–208.
- 122 Y. Kabri, A. Gellis and P. Vanelle, Eur. J. Org. Chem., 2009, 4059– 4066.
- 123 O. G. Schramm, T. Oeser, M. Kaiser, R. Brun and T. J. J. Müller, Synlett, 2008, 359, 362.
- 124 E. Milner, S. Gardner, J. Moon, K. Grauer, J. Auschwitz, I. Bathurst, D. Caridha, L. Gerena, M. Gettayacamin, J. Johnson, M. Kozar, P. Lee, S. Leed, Q. Li, W. McCalmont, V. Melendez, N. Roncal, R. Sciotti, B. Smith, J. Sousa, A. Tungtaeng, P. Wipf and G. Dow, J. Med. Chem., 2011, 54, 6277–6285.
- 125 A. Robin, F. Brown, N. Bahamontes-Rosa, B. Wu, E. Beitz, J. F. J. Kun and S. L. Flitsch, J. Med. Chem., 2007, 50, 4243–4249.
- 126 M. D'hooghe, S. Kenis, K. Vervisch, C. Lategan, P. J. Smith, K. Chibale and N. De Kimpe, Eur. J. Med. Chem., 2011, 46, 579–587.
- 127 C. Nguyen, G. F. Ruda, A. Schipani, G. Kasinathan, I. Leal, A. Musso-Buendia, M. Kaiser, R. Brun, L. M. Ruiz-Pérez, B.-L. Sahlberg, N. G. Johansson, D. González-Pacanowska and I. H. Gilbert, J. Med. Chem., 2006, 49, 4183–4195.
- 128 S. Urgaonkar, J. F. Cortese, R. H. Barker, M. Cromwell, A. E. Serrano, D. F. Wirth, J. Clardy and R. Mazitschek, Org. Lett., 2010, 12, 3998– 4001.
- 129 B. R. Shenai, P. S. Sijwali, A. Singh and P. J. Rosenthal, J. Biol. Chem, 2000, 275, 29000–29010.
- 130 M. Dua, P. Raphael, P. S. Sijwali, P. J. Rosenthal and M. Hanspal, Mol. Biochem. Parasitol., 2001, 116, 95–99.
- 131 R. Ettari, E. Nizi, M. E. Di Francesco, M.-A. Dude, G. Pradel, R. Vićík, T. Schirmeister, N. Micale, S. Grasso and M. Zappalà, J. Med. Chem., 2008, 51, 988–996.
- 132 R. Ettari, N. Micale, T. Schirmeister, C. Gelhaus, M. Leippe, E. Nizi, M. E. Di Francesco, S. Grasso and M. Zappalà, J. Med. Chem., 2009, 52, 2157–2160.
- 133 F. Bova, R. Ettari, N. Micale, C. Carnovale, T. Schirmeister, C. Gelhaus, M. Leippe, S. Grasso and M. Zappalà, Bioorg. Med. Chem., 2010, 18, 4928–4938.
- 134 D. Nöteberg, E. Hamelink, J. Hultén, M. Wahlgren, L. Vrang, B. Samuelsson and A. Hallberg, J. Med. Chem., 2003, 46, 734–746.
- 135 K. Ersmark, I. Feierberg, S. Bjelic, E. Hamelink, F. Hackett, M. J. Blackman, J. Hultén, B. Samuelsson, J. Åqvist and A. Hallberg, J. Med. Chem., 2004, 47, 110–122.
- 136 K. Ersmark, M. Nervall, E. Hamelink, L. K. Janka, J. C. Clemente, B. M. Dunn, M. J. Blackman, B. Samuelsson, J. Åqvist and A. Hallberg, J. Med. Chem., 2005, 48, 6090–6106.
- 137 D. Muthas, D. Nöteberg, Y. A. Sabnis, E. Hamelink, L. Vrang, B. Samuelsson, A. Karlén and A. Hallberg, Bioorg. Med. Chem., 2005, 13, 5371–5390.
- 138 S.-r. Choi, A. Pradhan, N. L. Hammond, A. G. Chittiboyina, B. L. Tekwani and M. A. Avery, J. Med. Chem., 2007, 50, 3841– 3850.
- 139 V. J. Bulbule, K. Rivas, C. L. M. J. Verlinde, W. C. Van Voorhis and M. H. Gelb, J. Med. Chem., 2008, 51, 384–387.
- 140 N. Bouloc, J. M. Large, E. Smiljanic, D. Whalley, K. H. Ansell, C. D. Edlin and J. S. Bryans, Bioorg. Med. Chem. Lett., 2008, 18, 5294–5298.
- 141 K. M. Orrling, M. R. Marzahn, H. Gutiérrez-de-Terán, J. Åqvist, B. M. Dunn and M. Larhed, Bioorg. Med. Chem., 2009, 17, 5933– 5949.
- 142 H. E. Howard-Lock, C. J. L. Lock, M. L. Martins, P. S. Smalley and R. A. Bell, Can. J. Chem., 1986, 64, 1215–1219.
- 143 D. Lavanchy, Liver Int., 2009, 29, 74–81.
- 144 J. F. Perz, G. L. Armstrong, L. A. Farrington, Y. J. F. Hutin and B. P. Bell, J. Hepatol., 2006, 45, 529–538.
- 145 C. Failla, L. Tomei and R. Defrancesco, J. Virol., 1994, 68, 3753–3760.
- 146 M. Nilsson, A. K. Belfrage, S. Lindstrom, H. Wahling, C. Lindquist, S. Ayesa, P. Kahnberg, M. Pelcman, K. Benkestock, T. Agback, L. Vrang, Y. Terelius, K. Wikstrom, E. Hamelink, C. Rydergard, M. Edlund, A. Eneroth, P. Raboisson, T.-I. Lin, H. de Kock, P. Wigerinck, K. Simmen, B. Samuelsson and Å. Rosenquist, Bioorg. Med. Chem. Lett., 2010, 20, 4004–4011. Downloaded by State At Albany on 24 March 2013 By State University of New York at Albany on 24 March 2013 By State University of New York at Albany on 24 March 2013 By State University of New York at Albany on 24 March 20
	- 147 A. Lampa, A. E. Ehrenberg, A. Vema, E. Åkerblom, G. Lindeberg, U. H. Danielson, A. Karlén and A. Sandström, Bioorg. Med. Chem., 2011, 19, 4917–4927.
	- 148 M. Pompei, M. E. Di Francesco, S. Pesci, U. Koch, S. E. Vignetti, M. Veneziano, P. Pace and V. Summa, Bioorg. Med. Chem. Lett., 2010, 20, 168–174.
	- 149 J. Gising, P. Örtqvist, A. Sandström and M. Larhed, Org. Biomol. Chem., 2009, 7, 2809–2815.
	- 150 P. Örtqvist, J. Gising, A. E. Ehrenberg, A. Vema, A. Borg, A. Karlén, M. Larhed, U. H. Danielson and A. Sandström, Bioorg. Med. Chem., 2010, 18, 6512–6525.
	- 151 X. Y. Wu, R. Rönn, T. Gossas and M. Larhed, J. Org. Chem., 2005, 70, 3094–3098.
	- 152 R. Rönn, A. Lampa, S. D. Peterson, T. Gossas, E. Åkerblom, U. H. Danielson, A. Karlén and A. Sandström, Bioorg. Med. Chem., 2008, 16, 2955–2967.
	- 153 R. Rönn and A. Sandström, Curr. Top. Med. Chem., 2008, 8, 533–562.
	- 154 J. M. Ontoria, E. H. Rydberg, S. Di Marco, L. Tomei, B. Attenni, S. Malancona, J. I. M. Hernando, N. Gennari, U. Koch, F. Narjes, M. Rowley, V. Summa, S. S. Carroll, D. B. Olsen, R. De Francesco, S. Altamura, G. Migliaccio and A. Carfi, J. Med. Chem., 2009, 52, 5217–5227.
	- 155 S. E. Lazerwith, G. Bahador, E. Canales, G. Cheng, L. Chong, M. O. Clarke, E. Doerffler, E. J. Eisenberg, J. Hayes, B. Lu, Q. Liu, M. Matles, M. Mertzman, M. L. Mitchell, P. Morganelli, B. P. Murray, M. Robinson, R. G. Strickley, M. Tessler, N. Tirunagari, J. Wang, Y. Wang, J. R. Zhang, X. Zheng, W. Zhong and W. J. Watkins, ACS Med. Chem. Lett., 2011, 2, 715–719.
	- 156 M. Larhed, G. Lindeberg and A. Hallberg, Tetrahedron Lett., 1996, 37, 8219–8222.
	- 157 A. Lampa, A. E. Ehrenberg, S. S. Gustafsson, A. Vema, E. Åkerblom, G. Lindeberg, A. Karlén, U. H. Danielson and A. Sandström, Bioorg. Med. Chem., 2010, 18, 5413–5424.
	- 158 P. Örtqvist, S. D. Peterson, E. Åkerblom, T. Gossas, Y. A. Sabnis, R. Fransson, G. Lindeberg, U. H. Danielson, A. Karlén and A. Sandström, Bioorg. Med. Chem., 2007, 15, 1448–1474.
	- 159 J. Q. Wan, Y. Xia, Y. Liu, M. H. Wang, P. Rocchi, J. H. Yao, F. Q. Qu, J. Neyts, J. L. Iovanna and L. Peng, J. Med. Chem., 2009, 52, 1144– 1155.
	- 160 R. Z. Zhu, M. H. Wang, Y. Xia, F. Q. Qu, J. Neyts and L. Peng, Bioorg. Med. Chem. Lett., 2008, 18, 3321–3327.
	- 161 T. D. Tran, D. C. Pryde, P. Jones, F. M. Adam, N. Benson, G. Bish, F. Calo, G. Ciaramella, R. Dixon, J. Duckworth, D. N. A. Fox, D. A. Hay, J. Hitchin, N. Horscroft, M. Howard, I. Gardner, H. M. Jones, C. Laxton, T. Parkinson, G. Parsons, K. Proctor, M. C. Smith, N. Smith and A. Thomas, Bioorg. Med. Chem. Lett., 2011, 21, 2389–2393.
	- 162 A. Gopalsamy, K. Lim, G. Ciszewski, K. Park, J. W. Ellingboe, J. Bloom, S. Insaf, J. Upeslacis, T. S. Mansour, G. Krislinamurthy, M. Damarla, Y. Pyatski, D. Ho, A. Y. M. Howe, M. Orlowski, B. Feld and J. O'Connell, J. Med. Chem., 2004, 47, 6603–6608.
	- 163 E. I. Pécheur, D. Lavillette, F. Alcaras, J. Molle, Y. S. Boriskin, M. Roberts, F. L. Cosset and S. J. Polyak, Biochemistry, 2007, 46, 6050–6059.
	- 164 G. Sellitto, A. Faruolo, P. de Caprariis, S. Altamura, G. Paonessa and G. Ciliberto, Bioorg. Med. Chem., 2010, 18, 6143–6148.